



Original Article



Prognostic value of neutrophil CD64 in low birth weight neonates with sepsis

Madiha A. Sayed^{1*}, Amal M.Kamal², Mohamed A. Abdelhakim², Reham R. Hassan²

DOI: 10.21608/ANJ.2019.45815

*Correspondence: Pediatric Department, Faculty of Medicine, Minia university, Egypt

Email: madialy1970@yahoo.com@yahoo.com

[Full list of author information is available at the end of the article.](#)

Abstract

Background: Neonatal sepsis represents a major health problem with high mortality and morbidity rates. Although early diagnosis of neonatal sepsis is very important for proper management yet it remains a difficult task. Neutrophil CD64 (nCD64) is used as a marker for the diagnosis of sepsis, requiring a small sample volume, short turnaround time.

Objective: In this study we aimed to study the diagnostic performance of nCD64 against routine markers in low birth weight neonates (LBWN) with sepsis.

Methods: A case control study was conducted on 40 LBWN suspected clinically to have early onset neonatal sepsis against 20 neonates clinically free of sepsis as control. Investigations included CBC, CRP, blood culture and nCD64 expression.

Results: among the studied markers of sepsis; immature neutrophil count, immature /mature ratio, immature/total ratio, CRP and nCD64 were significantly higher in suspected group than control (p value 0.007, 0.001, 0.002, 0.001, and 0.001 respectively). Among the group of neonates with suspected sepsis, blood culture of 11 cases (27.5%) did not show growth. nCD64 showed the highest sensitivity and specificity; 100% each. Immature neutrophil count and total leucocytic count showed the lowest sensitivity 40% and mature neutrophil showed the lowest specificity 45%. The expression of nCD64 in those neonates who died as a complication of sepsis was significantly higher than those who survived (p value 0.001).

Conclusion: nCD64 is a reliable marker for the diagnosis of early onset neonatal sepsis in LBWN with a significant predictive value for disease course.

Key words: Low birth weight neonates; Neonatal sepsis; Neutrophil CD64.

Introduction

Early onset neonatal sepsis (EONS) is a life-threatening condition for neonates during their first 72 hours of life. In general, neonatal sepsis, sepsis neonatorum or neonatal septicemia are synonyms that being used to describe a systemic response of a neonate to infection [1]. Multiple maternal, neonatal and environmental factors play a role in the development of neonatal sepsis [2].

The associated risk factors for EONS include: preterm delivery, premature rupture of membranes (PROM), maternal urinary tract infection (UTI), maternal fever, maternal chorioamnionitis, group B streptococcal (GBS) infections rectovaginal colonization, foul smelling liquor, multiple per vaginum examinations, difficult or prolonged labour, aspiration of meconium, very low birth weight (VLBW), prematurity, asphyxia, low Apgar score and male sex [3]. The micro-organisms most commonly associated with EONS are GBS, E.coli,

haemophilus influenzae and listeria monocytogenes [4].

In Egypt, rates of neonatal sepsis (EONS) exceeding 50% especially in neonatal intensive care unit (NICU) with a mortality rate of 51% for proven EONS and 42.9% for proven LONS [5, 6, 7]. The lack of a well-established laboratory marker for an early diagnosis of neonatal sepsis increases the challenge for management as rapid definitive diagnosis is required. Although blood culture is the gold standard technique it has a long turnaround time (TAT) in addition to its poor positive and negative predictive values [8]. On the other hand, white blood cell (WBC) counts, absolute neutrophil count (ANC), immature neutrophil count, the ratio of immature: Total neutrophils (I:T) and immature : mature neutrophils (I:M) are commonly used parameters as screening tests for the diagnosis of neonatal sepsis, [9, 10] yet they have poor positive predictive value (PPV) and poor diagnostic accuracy in terms of sensitivity and specificity [4].

Cytokines can be used for screening of sepsis yet they have poor specificity and their levels are linked to the immune status of the neonate [11]. Inflammatory markers as C-reactive protein (CRP) and procalcitonin (PCT) are commonly used as routine markers for the diagnosis of sepsis. However, CRP requires 6-8 hours for being synthesized after stimulation and 24 hours to reach the peak. Its quantitative assay has no superiority over WBC counts or ratios except in monitoring the effect of treatment if measured serially [12]. Although it is more accurate than CRP for the diagnosis of neonatal sepsis, procalcitonin has moderate accuracy for the diagnosis of neonatal sepsis [13].

Neutrophils CD64 (nCD64) is a surface marker expressed in a very low concentration in resting conditions however its level increases 5-10 folds after infectious stimulations and the level is correlated with the process of phagocytosis [14]. It is considered a sensitive laboratory marker for

diagnosing neonatal sepsis. It is superior to CRP as it is activated even before CRP starts to rise [15]. The assessment of nCD64 is relatively simple and fast; it requires a small blood volume with no special precautions and no effect of previous antibiotic use [16]. In this study we aimed to evaluate the nCD64 for early diagnosis of EONS against other markers and to evaluate its value in predicting disease course in LBWN.

Methods

This prospective study was conducted in the Neonatal intensive care unit& Department of Clinical Pathology, at Minia University Hospital for Obstetrics and children. Written informed consent was signed by the parents of neonates enrolled in the study. The protocol of the study follows the principles outlined in the Declaration of Helsinki at World Medical Association. (<https://www.wma.net/policies-post/wma-declaration-of-helsinki-ethical-principles-for-medical-research-involving-human-subjects>).

Low birth weight neonates, who were admitted to the NICU were included in the study during the period from June 2016 to January 2017. They were classified into the following two groups; group I included 40 neonates having three or more risk factors for EONS addition to strong clinical suspicion of sepsis. Their gestational age ranged from (35-40 wks.), their weights ranged from (1.4 – 2.5 kg, mean \pm SD of 2.16 ± 0.28), 26 were males and 14 females, this group was subdivided into two subgroups; those who survived which included 22 neonates and the other for those who died as a complication of sepsis and included 18 neonates. A control group, group II, included 20 apparently healthy neonates with matched gestational age whom blood samples were taken for other routine investigations as ABO grouping and thyroid functions.

Ethylenediaminetetraacetic acid (EDTA) samples were used for hematological studies on Celtac Es, NIHON KOHDEN COPORATION, AUTOMATED

HEMATOLOGY ANALYSER, Japan and for flowcytometric study of n-CD64 expression using BD FACS cantotm II USA according to the following protocol: One hundred μ l of EDTA blood were used for the evaluation of nCD64 expression. For each sample, 2 tubes were labeled, one for fluorescein isothiocyanate (FITC) mouse anti-human CD64 monoclonal antibody (BD-Bioscience), the other tube for negative isotypic control (FITC) Mouse IgG1 κ Isotype control. Fifty μ l of samples were delivered in each tube. Four μ l of monoclonal antibodies were added to respective tubes. Then both tubes were vortexed, incubated for 15 minutes.

Three ml of lysing solution was added to each tube then the tubes were vortexed and incubated for just 10 minutes followed by centrifugation. The supernatant was discarded and phosphate buffered saline (PBS) was added to the sediment and mixed thoroughly, then centrifuged. The supernatant was discarded and cells were suspended in

300 µl PBS to be ready for acquiring data by the flowcytometric analysis. In flowcytometry, cell surface expression of nCD64 was determined at 468 nm wavelength laser excitation and the emitted fluorescence was monitored with a detector optimized to collect peak emissions at 504 – 541 nm. Neutrophils phenotyping was done by gating according to forward scatter (size) and side scatter (granularity) strategy. Results were expressed molecules of equivalent soluble fluorochrome (MESF). Serum samples were used for CRP assay quantitatively using Human C-Reactive Protein (CRP) ELISA Kit, the Cell Biolabs, Inc. San Diego, CA 92126. USA.

Ethical considerations

The study was revised and approved by the scientific committee of the pediatrics department, Minia University. Written and verbal consent was obtained from the parents of babies prior to inclusion in the study.

Statistical analysis

The data were encoded, entered and processed on computer using Graph Pad prism 4. Data were presented as mean ± standard deviation (SD). Statistical analysis was carried out using paired sample t test and Mann–Whitney test. Positive predictive value, negative predictive value, sensitivity, and specificity were obtained using optimal cutoff levels. Correlations were calculated by the Pearson and Spearman rank methods. Probability values <0.05 were considered to be significant.

Results

Table (1) showed that the total leucocytic count in the group with suspected sepsis ranged from 10000 - 32300 cell/mm³ with a mean of 16600 and SD ±5800, while the number of TLC count in the control group ranged from 10000 to 25000cell/ mm³ with a mean of 17700 and SD ±4400. There was no statistically significant difference between the two groups regarding to the TLC count (p-value 0.190). The absolute neutrophil count (ANC) count in group with

suspected sepsis ranged from 4900 to 16907 cell/mm³ with a mean of 6019.7 and SD ±1781, while the number of ANC in control group ranged from 5705 - 12152 cell/mm³ with a mean of 7542.5 and SD ±1695. There was no statistically significant difference between the two groups regarding to the ANC (p-value 0.371).

The count of mature neutrophil in group with suspected sepsis ranged from 3220 to 13398 cell /mm³ with a mean of 7985.4 and SD ±2216, while the number of mature neutrophil count in control group ranged from 3912 to 10388 cell/mm³ with a mean of 6019.7 and SD ±1520. There was no statistically significant difference between the two groups regarding to the mature neutrophil count (p-value 0.987).

The count of immature neutrophil (bands, metamyelocytes, myelocytes and promyelocytes) in group with suspected sepsis ranged from 1144 to 3876 cell/mm³ with a mean of 1965.7 and SD ±618, while the number of immature

neutrophil count in control group ranged from 900 to 2000 cell/mm³ with a mean of 1522.9 and SD ±318.8. Immature neutrophil count was statistically significant low in group with suspected sepsis when compared to control group (p-value 0.007). The I/M ratio in group with suspected sepsis ranged from 0.15 to 1.5 with a mean of 0.34 and SD ±0.2, while I/M ratio in control group ranged from 0.1 to 0.4 with a mean of 0.24 and SD ±0.07. I/M ratio were statistically significant higher in group with suspected sepsis when compared to control group (p-value 0.001).

The I/T ratio in group with suspected sepsis ranged from 0.13 to 0.8 with a mean of 0.26 and SD ±0.1, while I/T ratio in control group ranged from 0.14 to 0.3 with a mean of 0.2 and SD ±0.04. I/T ratio was statistically significant higher in group with suspected sepsis when compared to control group (p-value 0.001). Among the studied group of neonates with suspected sepsis, blood culture results revealed that nine cases

(22.5%) had staphylococcus epidermis; seven cases (17.5%) had staphylococcus sapropheticus; five cases (12.5%) had klebsiella pneumoniae; two cases (5%) had pseudomonas species; two cases (5%) had acintobacter species; two cases (5%) had enterobacter species; one case (2.5%) had streptococcus pyogenes and one case (2.5%) had candida species, while eleven cases (27.5%) showed no growth as shown in figure(1).

CRP in group with suspected sepsis ranged from 5.5 to 68 mg/l with a mean of 20.39 and SD \pm 15.07, while the CRP in control group ranged from 1.1 to 18.5 mg/l with a mean of 5.4 and SD \pm 5.05. The CRP level was statistically significant high in group with suspected sepsis group when compared to control group (p-value 0.0001) nCD64 in group with suspected sepsis ranged from 2006 to 5192 MESF with a mean of 2622.4 and SD \pm 784.7, while the nCD64 in control group ranged from 625 to 1023 MESF with a mean of 803.9 and SD \pm 140. nCD64 level was statistically

significant low in group with suspected sepsis when compared to control group (p-value 0.0001) as shown in figure-2.

Table-2 and figures 3-6 showed a receiver operating characteristic (ROC) curves were generated to calculate the sensitivity, specificity, positive predictive value (PPV) and negative predictive value (NPV) using optimal cutoff values for the studied parameters. A comparison of each test showed that neutrophil CD64 a sensitivity of 100%, a specificity of 100%, PPV 100% and NPV of 100% (figure 3-a). CRP showed a sensitivity of 95.2%, a specificity of 83.3%; PPV 90.91% and NPV 90.91% (figure 3-b). Blood culture showed a sensitivity of 77.5%, a specificity of 50%, PPV of 76% and NPV of 53%. Absolute neutrophil count showed a sensitivity of 72.5%, a specificity of 55%, PPV of 76.3% and NPV 50%. I/M ratio showed a sensitivity of 72%, a specificity of 85%, PPV of 90% and NPV of 60%, I/T Ratio showed a sensitivity of 72%, a specificity of 45%, PPV of 71% and NPV of 54%.

Mature neutrophil showed a sensitivity of 70%, a specificity of 45%, PPV of 71% and NPV of 42%, Immature neutrophil showed a sensitivity of 40%, a specificity of 100%, PPV of 100% and NPV of 25%, TLC showed a sensitivity of 40%, a specificity of 85%, PPV 84% and NPV 40%.

Table (3) showed that among the studied 40 neonates with sepsis, eighteen patients died while twenty-two patients survived, the nCD64 expression level among those who died ranged between 2082 to 5192 MESF with a mean of 3160 and SD \pm 881-9, while in those who survived it ranged between 2006 to 2813 MESF with a mean of 2182.5 and SD \pm 251.6. NCD64 expression was statistically higher in patient who died than those who survived (P- value 0.001).

Discussion

Despite the increased awareness of infection control measures, introduction of potent antimicrobials and improvement of laboratory techniques, neonatal sepsis remains a global health

problem due to its significant contribution to high morbidity and mortality [17]. Early diagnosis of neonatal sepsis is a matter of clinical dilemma because of the overlapping clinical presentation [18]. There is a need for a sensitive specific test with a short TAT that would allow a safe cessation of antibiotics in neonates without infection and would recommend antibiotics for those with probable neonatal sepsis. Among 40 LBWN suspected to have sepsis, nCD-64 was estimated as an early marker of sepsis and compared to CRP, WBC counts and blood culture. Levels of these parameters were compared to those of a control group had 20 neonates without a single marker of sepsis.

Unlike most of the reports regarding EONS commonest pathogens (which are gram negative organisms representing maternal flora) [19-21]. We had more gram-positive pathogens as the leading cause for EONS (47.5%) and staphylococcus epidermidis being the commonest isolated organism (27.5%).

Preterm neonates included in this group may explain this discrepancy where staphylococcus epidermidis is the most common species of CONS associated with neonatal sepsis in preterm infants, which accounts for 60 to 93% of CONS bloodstream infections [22]. Yet, these results were in accordance with Sobaih and Al-Mandeel [8].

False-negative blood cultures in apparently septic neonates can be interpreted by poor timing or inadequate blood sample size, fastidious organism and maternal intake of antibiotics [23]. Blood culture is the gold standard laboratory technique for the diagnosis of early onset neonatal sepsis although the relatively long TAT (2-4 days), the inappropriate sensitivity in detecting bacteremia owing to the dilution of a relatively small sample, the transient bacteremia and the effect of previously administered antibiotics [24].

In the present study, hematological laboratory indices were estimated among cases and control. No significant

association was found between total leukocyte counts and neonatal septicemia. Similar result was obtained in a study done by Mayuga and Isleta [25]. In a study by Ottolini et al., it was found that TLC are of limited value in the diagnosis of septicemia in newborns [26]. Total leucocytic counts are particularly unreliable indicator of infection during the first several hours of early-onset (within 48 h of birth) sepsis because their high values are initially normal [27]. The inadequate specificity of mature neutrophil count and the poor sensitivity of immature neutrophil count as markers of EONS were reported previously. During sepsis, a 'left shift' of neutrophils happens because of immature neutrophils released from marrow which increases the ratio of immature to total neutrophils [28]. Our results revealed that the I/T ratio, I/M ratio of neutrophils and immature neutrophils are higher in septic neonates compared to normal neonates. These results were in agreement with Mondal et al., [29] who

found that the hematologic profiles of neonates with septicemia were characterized by higher I/T ratio, Bhandari et al., found that the hematologic profiles of neonates with septicemia were characterized by higher ANC, I/T ratio and immature neutrophil [30], Moreover, TLC and differential counts lacks the proper specificity as their automatic assessment is affected by the presence of nucleate red blood cells while manual count and blood film examination requires special skills [31]. This explains the inadequate sensitivity and specificity obtained in this work for blood culture as a marker of sepsis in LBWN.

CRP, a peptide synthesized by the liver in response to infection or inflammatory processes [32]. Our result revealed that CRP was statistically significant in septic neonates compared to control; this was in accordance with other researches [12, 33, 34]. However, a positive CRP result does not differentiate between systemic inflammatory response and sepsis,

neither between bacterial infection and non-bacterial infections [35]. Moreover, the latency between infection and synthesis of CRP till reaching the peak level affects the sensitivity and specificity as reported in this work [36].

The high affinity antibody receptor CD64 is expressed at a very low level on the surface of neutrophils in the absence of an infection. The expression of CD64 on activated neutrophils markedly increases after an episode of bacterial infection [37]. The results of this study showed a significantly higher expression level of nCD64 in LBWNs with suspected sepsis when compared to control as reported previously [38,39]. Shi and his colleagues reported in a meta-analysis a lower pooled sensitivity and specificity for nCD64 in neonatal sepsis than other markers [40], this disagrees with our results. The higher frequency gram positive organisms in this work are accompanied by higher expression of neutrophils in addition to the selection LBWN as candidates may be a reason for

this discrepancy in results. The presence of high sensitivity and specificity of CD64 in our study, high positive and negative predictive values of the test make it of a great value in diagnosing neonatal sepsis. More over upregulation of CD64 in the group of neonates who died added prognostic importance.

Until now, there is no reliable marker that can be used alone to predict the outcome of neonatal sepsis, yet our work may provide preliminary results for a single marker that can predict disease outcome in LBWN in environment with high prevalence of gram-positive organisms. Further studies on a larger scale at different environments are required.

Conclusions

nCD64 is a reliable marker for the diagnosis of early onset neonatal sepsis in LBWN with a significant predictive value for disease course.

Acknowledgements

To all NICU staff members Minia University Hospital for Obstetrics and children

Author's contributions

SS and EA conceived the study. ME revised the patients' medical reports and the final manuscript. All authors revised the final draft of the manuscript

Conflict of interest

The authors have no conflict of interests to declare.

Funding

This study received no special funding and was totally funded by the authors.

Author's details

¹Pediatric Department, Faculty of Medicine, Minia University, Egypt

²Clinical-Pathology Department, Faculty of Medicine, Minia University, Egypt

Date received: 13th December, 2018, accepted 27th January, 2019

References

1. Gotoff SP.: Infections of the neonatal infant. In "Nelson textbook of pediatrics", 16th Ed (Behrman RE Kliegman RM Jenson HB, eds). WB Saunders Philadelphia, USA (2000):538-552.
2. Kardana IM.: Incidence and factors associated with mortality of neonatal sepsis. Paediatr Indones (2011); 51 (3):144-148.
3. Chacko B and Sohi I.: "Early onset neonatal sepsis," Indian Journal of Pediatrics (2005); 72 (1):23-26

4. Tripathi S and Malik GK.: Neonatal Sepsis: past, present and future; a review article. *Internet Journal of Medical Update* (2010); 5 (2): 45-54.
5. Shehab El-Din EM, El-Sokkary MM, Bassiouny MR and Hassan R.: Epidemiology of Neonatal Sepsis and Implicated Pathogens: A Study from Egypt. *Biomed Research International* (2015); Volume 2015, Article ID 509484, 11 pages .<http://dx.doi.org/10.1155/2015/509484>
6. Moore KL, Kainer MA, Badrawi N, Afifi S, Wasfy M, Bashir M, Jarvis WR, Graham TW, el-Kholy A, Gipson R, Jernigan DB and Mahoney F.: "Neonatal sepsis in Egypt associated with bacterial contamination of glucose-containing intravenous fluids," *Pediatric Infectious Disease Journal* (2005); 24(7):590–594
7. El-Shiekh H, Gaafar M, Yosri M, Hassan D and Said H.: Study of Bacteria Causing Septicemia in Neonatal Intensive Care Unit. *Egyptian Journal of Medical Microbiology* (2016); 25 (1):37-44.
8. Paolucci M, Landini MP and Sambri V.: How can the microbiologist help in diagnosing neonatal sepsis? *International Journal of Pediatrics* (2012); doi: 10.1155/2012/120139
9. Polin RA.: Management of neonates with suspected or proven early-onset bacterial sepsis. *Pediatrics*; 129(5): 1006-1015. Murphy K and Weiner J. (2012): Use of leukocyte counts in evaluation of early-onset neonatal sepsis. *Pediatr Infect Dis J* (2012); 31(1): 16-19.
10. Murphy K and Weiner J.: Use of leukocyte counts in evaluation of early-onset neonatal sepsis. *Pediatr Infect Dis J*(2012); 31(1): 16-19.
11. Arunachalam AR and Pammi M.: Biomarkers in early-Onset Neonatal Sepsis: An Update. *Ann Clin Med Microbio*(2015); 1(2): 1007.
12. Chirico G and Loda C.: Laboratory aid to the diagnosis and therapy of infection in the neonate. *Pediatric Reports* (2011), 3(1):34-42
13. Yu Z1, Liu J, Sun Q, Qiu Y, Han S and Guo X.: The accuracy of the procalcitonin test for the diagnosis of neonatal sepsis: a meta-analysis. *Scand J Infect Dis* (2010). 42(10):723-33
14. Song SH, Kim HK, Park MH and Cho HI.: Neutrophil CD64 expression is associated with severity and prognosis of disseminated intravascular coagulation. *Thromb Res* (2008); 121(4):499-507
15. Mally P, Xu J and Hendricks-Muñoz KD.: Biomarkers for neonatal sepsis: recent developments. *Research and Reports in Neonatology* (2014); 4:157–16.
16. Hoffmann J.: Neutrophil CD64: A diagnostic marker for infection and sepsis. *Clin. Chem. Lab. Med.*; 47(8):903-916.
17. Naher H and Khamael A. (2013): Neonatal Sepsis; The Bacterial Causes and the Risk Factors. *Int. Res. J. Medical Sci.* (2009); 1(6): 19-22.
18. Dhlamini M, Suchard M and Wiggill T.: Neutrophil CD64 has a high negative predictive

- value for exclusion of neonatal sepsis. *S. Afr. J. CH.* (2013);7(1):25-29.
19. Fahmey SS.: Early-onset sepsis in a neonatal intensive care unit in BeniSuef, Egypt: bacterial isolates and antibiotic resistance pattern, *Korean J Pediatr* (2013); 56(8):332-337
20. Kilani RA and Basamad M.: Pattern of proven bacterial sepsis in a neonatal intensive care unit in Riyadh-Saudi Arabia: a 2-year analysis. *J Med Liban* (2000); 48: 77-83.
21. Panwar C, Kaushik SL, Kaushik R and Sood A.: Correlation of neonatal and maternal clinico-hematological parameters as predictors of early onset neonatal sepsis, *International journal of temporary*, vol 4(2017); no 1.
22. Simonsen KA, Anderson-Berry AL, Delair, SF and Davies HD.: Early-Onset Neonatal Sepsis. *Clinical Microbiology Reviews* (2014), 27(1), 21–47.
23. Sobaih BH and Al-Mandeel H.: Early and Late Onset Neonatal Sepsis in Very Low Birth Weight Infants in a Tertiary Center in Saudi Arabia, *J Neonatal Biol*(2014), 3:5 PP. 2-4.
24. Mishra U K, Jacobs S E, Doyle L W, and Garland S M: Newer approaches to the diagnosis of early onset neonatal sepsis. *Arch Dis Child Fetal Neonatal Ed* (2006); 91(3): F208–F212.
25. Mayuga WAB and Isleta PFD: Clinical correlation of neonatal and maternal hematological parameters as predictors of neonatal sepsis. *PIDSP J.* (2005); 9:36–43.
26. Ottolini MC, Lundgren K, Mirkinson LJ, Cason S and Ottolini MG.: Utility of complete blood count and blood culture screening to diagnose neonatal sepsis in the asymptomatic at risk newborn. *Pediatr Infect Dis J.* (2003); 22:430–434.
27. Zhou B , Liu X , Wu JB , Jin B , and Zhang YY: Clinical and microbiological profile of babies born with risk of neonatal sepsis. *ExpTher Med* (2016); 12(6): 3621–3625.
28. Li W, Wu AH, Zhu S, Li J, Wu R and D'Angelo J.: EGCG induces G-CSF expression and neutrophilia in experimental sepsis. *Immunol Res* (2015). 63 (1-3):144-52
29. Mondal S, Dipanwita R and Bandyopadhyay R: Neonatal sepsis: role of a battery of immunohematological tests in early diagnosis. *International J. of Applied and Basic Medical Research.* (2012); 2(1):43-47.
30. Bhandari V, Wang C, Rinder C and Rinder H.: Hematologic profile of sepsis in neonates: neutrophil CD64 as a diagnostic marker. *Pediatrics* (2008); 121(1):129-34.
31. Ng P and Lam H.: Diagnostic markers for neonatal sepsis. *Curr.Opin.Pediatr.* (2006); 18(2): 125-131
32. Markanday A.: Acute Phase Reactants in Infections: Evidence-Based Review and a Guide for Clinicians. *Open Forum Infectious Diseases* (2015), 2(3), 98.
33. Chacha F, Mirambo MM, Mushi MF, Kayange N, Zuechner A and Kidenya BR.: Utility of qualitative C- reactive protein assay and white blood cells counts in the diagnosis of neonatal septicaemia at Bugando Medical Centre,

- Tanzania. *BMC Pediatr*3 (2014); 14:248.. doi: 10.1186/1471-2431-14-248.
34. Hedegaard SS, Wisborg K and Hvas AM.:Diagnostic utility of biomarkers for neonatal sepsis—a systematic review. *Infect Dis (Lond)* (2014). 2015; 47(3):117–24.
35. Krishnavenil P and Gowda V.: Serum Amyloid A Protein levels in Neonatal Sepsis. *International Journal of Clinical Biochemistry and Research* (2017); 4(1):47-55.
36. QuJ, Lü X, Liu Y, and Wang X: Evaluation of procalcitonin, C-reactive protein, interleukin-6
37. Shah BA and Padbury JF.: Neonatal sepsis: An old problem with new insights. *Virulence* (2014), 5(1), 170–178.
38. Khalifa R, Shehata I and Elsayed M.: Diagnostic value of Neutrophil CD64 in patients with Systemic Inflammatory Immune Syndrome. *Egypt J. Med. Lab. Sci.* (2007); 16(1):1-13.
39. Mahmoud FM, Darwish NM, Hassan RA and Abou Shady NM.: Evaluation of CD64 detection on neutrophils and TLR-2 on monocytes by flowcutometry as markers for early diagnosis of Neonatal Sepsis. *International Journal of Advanced Research* (2014), Volume 2, Issue 7, 1235-1247.
40. Shi J, Tang J and Chen D: Meta-analysis of diagnostic accuracy of neutrophil CD64 for neonatal sepsis. *Ital J Pediatr* (2016); 42: 57.

Table 1: Comparison between the two groups regarding the leucocyte counts,, I/M and I/T ratios.

Variable	Cases N = 40	Controls N = 20	P –value
TLC (cell/mm³)			
Range	10000– 32300	10000- 25000	0.190
Mean ± SD	16600 ± 5800	17700 ± 4400	NS
Absolute neutrophil (cell/mm³)			
Range	4900- 16907	5705- 12152	0.371
Mean ± SD	6019.7 ± 1781	7542.5 ± 1695	NS
Mature neutrophil (cell/ mm³)			
Range	3220– 13398	3912- 10388	0.987
Mean ± SD	7985.4 ± 2216	6019.7 ± 1520	NS
Immature neutrophil (cell/ mm³)			
Range	1144- 3876	900 - 2000	0.007*
Mean ± SD	1965.7 ± 618	1522.9 ± 318.8	
I/M			
Range	0.15–1.5	0.10–0.40	0.001*
Mean ± SD	0.34 ± 0.20	0.24 ± 0.07	
I/T			
Range	0.13–0.80	0.14–0.30	0.002*
Mean ± SD	0.26 ± 0.10	0.20 ± 0.04	

I/M=immature neutrophils/Mature neutrophils

I/T= immature neutrophils/Total leucocytic count

Table 2: Sensitivity, Specificity, PPV, and NPV for markers of EONS in LBWN

Variable	Cut of value	Sensitivity	Specificity	Positive predictive value	Negative predictive value
nCD64	>1515	100%	100%	100%	100%
CRP	>6	95.2%	83.3%	90.91%	90.91%
Blood culture	---	77.5%	50%	76%	53%
TLC	>13.5	40%	85%	84%	41%
Absolute neutrophil	>6845	72.5%	55%	76.3%	50%
Mature neutrophil	>5160	70%	45%	71%	42%
Immature neutrophil	>2000	40%	100%	100%	45%
I/M Ratio	>0.28	72%	85%	90%	60%
I/T Ratio	>0.21	72%	80%	80%	54%

Table 3: Comparison between n-cd64 in neonates in relation to disease outcome

CD64	Died (n=18)	Survived (n=22)	P- value
Range	2082–5192	2006–2813	0.001*
Mean ± SD	3160 ± 881.9	2182.5 ± 251.6	

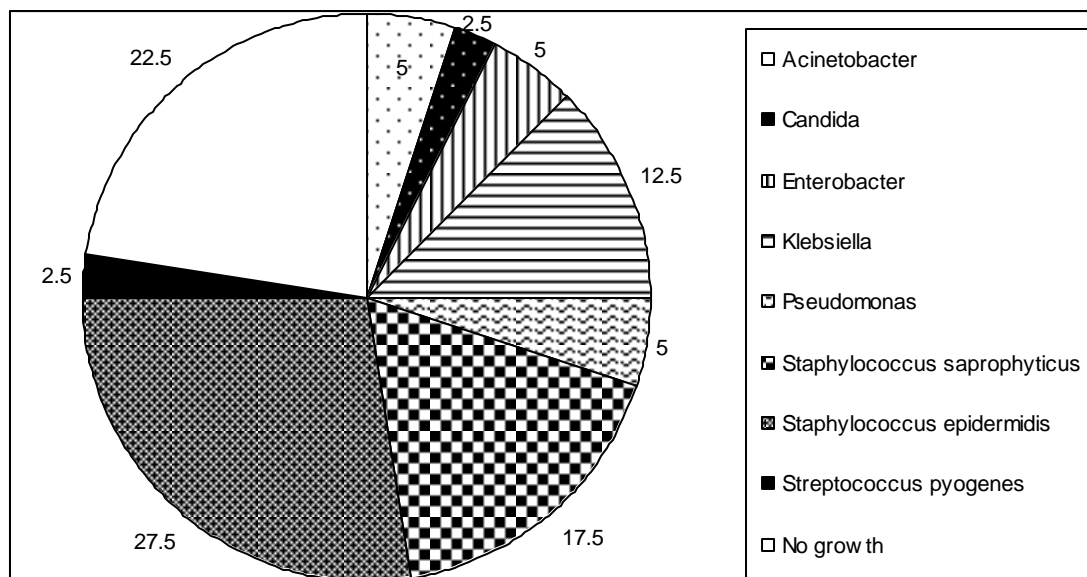


Figure 1: Frequency of organisms among the cases

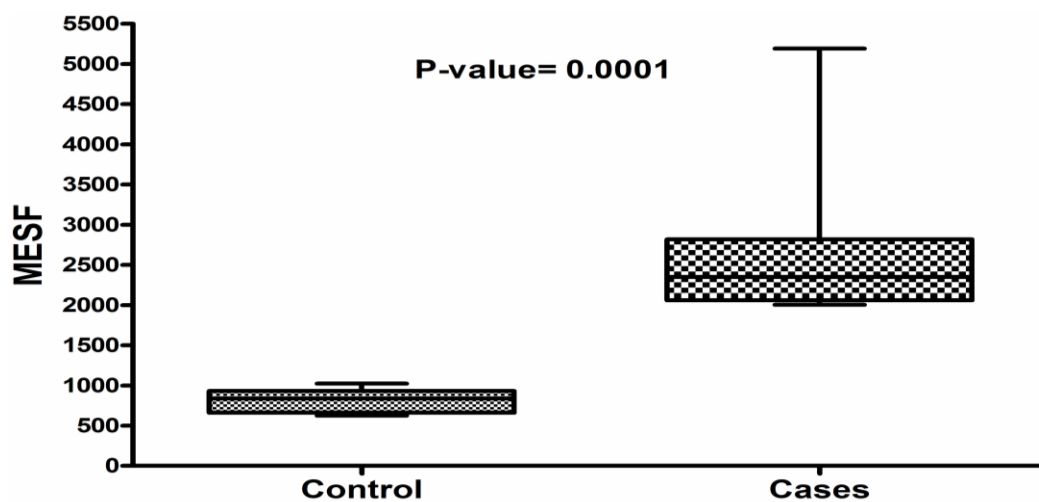


Figure 2: Comparison between the two groups regarding the nCD 64 expression level

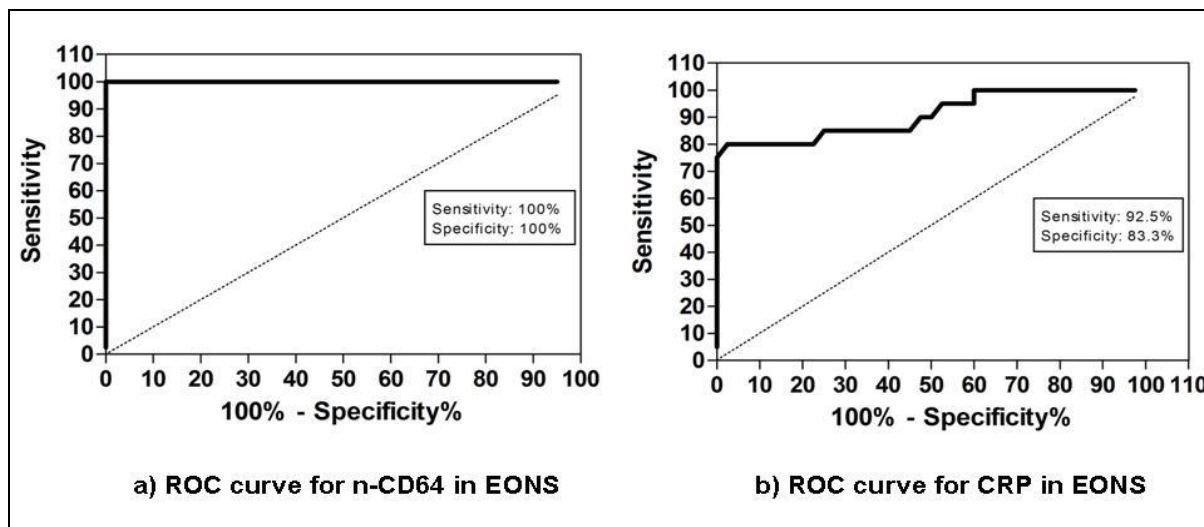


Figure 3: ROC curve for n-CD64 and CRP in EONS

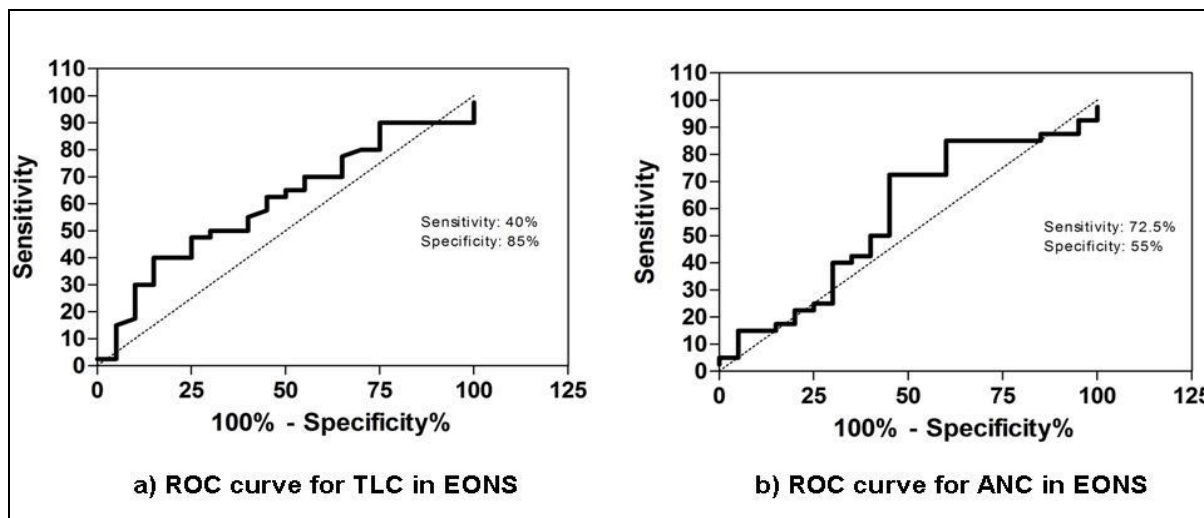


Figure 4: ROC curve for TLC and ANC in EONS

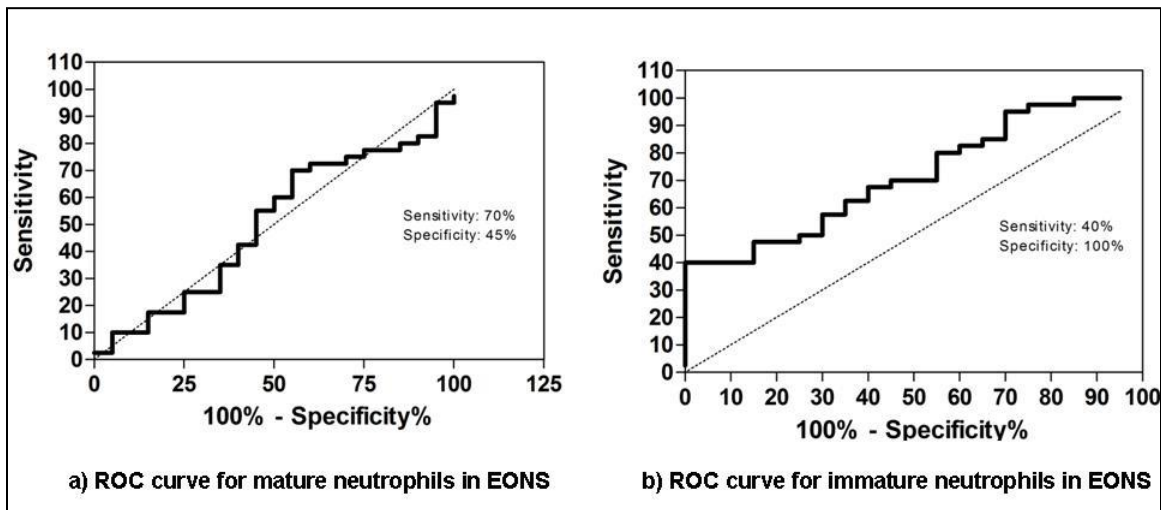


Figure 5: ROC curve for mature and immature neutrophils in EONS

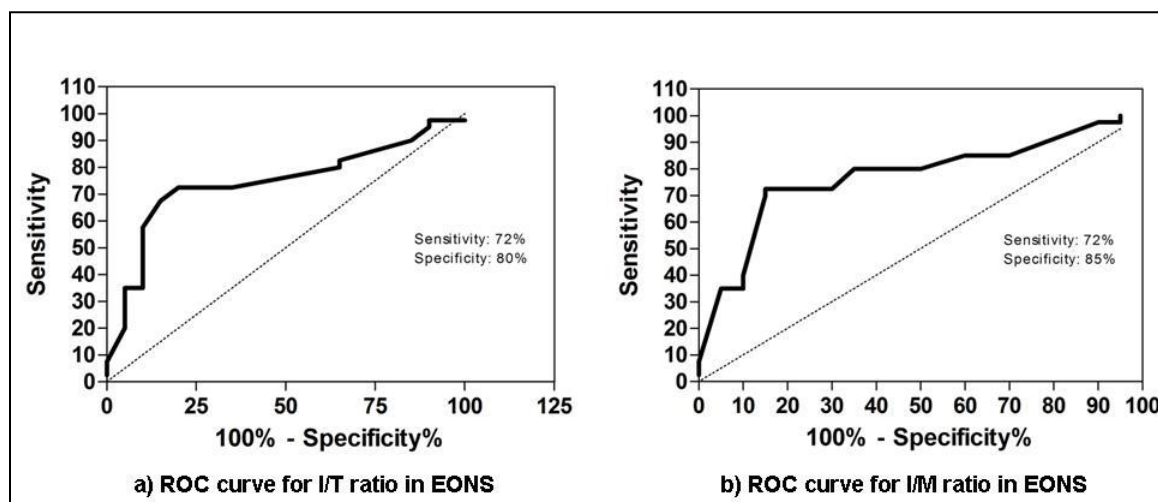


Figure 6: ROC curve for I/T and I/M ratios in EONS

Submit your next manuscript to **Annals of Neonatology Journal** and take full advantage of:

- Convenient online submission
- Thorough and rapid peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- No limit as regards tables or figures.
- Open Access research freely available for redistribution

Submit your manuscript at:

www.anj.journals.ekb.eg

Citation: Madiha A. Sayed; Amal M. Kamal; Mohammed A. Abdel Hakim; Reham R. Hassan. "Prognostic Value of Neutrophil CD 64 in Low Birth Weight Neonates with Sepsis". *Annals of Neonatology Journal*, 1, 1, 2019, 13-25. doi: 10.21608/anj.2019.45818



Copyright: Sayed et al. 2019. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC-BY) license (4).